

ARTUR PASTERNAK^{1, 2}, MIROSLAW SZURA², MAŁGORZATA MAZUR¹, IZABELA MRÓZ¹,
MACIEJ MATYJA³, ANDRZEJ MATYJA²

NUMBER AND DISTRIBUTION OF INTERSTITIAL CELLS OF CAJAL IN HUMAN GALLBLADDER

Abstract: Based on the review of current literature and on their own studies authors postulate that decreased number of interstitial cells in the wall of gallbladder may be pathognomonic for gallstone disease (cholelithiasis).

Key words: interstitial cells of Cajal, human gallbladder.

INTRODUCTION

It was Santiago Ramon y Cajal who described for the first time in the world interstitial cells [1, 2]. Following his opinion these cells played role of separate nervous cells, relatively rich within intestinal smooth muscle, where they united with autonomic nerve bundles forming dense. The cells varied according to shape. Some were fusiform, triangular, sometimes multipolar (star-like). In Cajal's opinion they could play modulatory role in contraction of smooth myocytes of gastrointestinal system [2].

Interstitial cells were detected in many other tissues, always in the vicinity of smooth muscle cells. ICC were identified in gastrointestinal tract of many species, i.e. mice [3], rats [4], guinea pigs [5]. In humans ICC were found in the wall of alimentary tract [6], pancreas [7], in the muscle of atria and ventricles [8–10], vagina [11], mammary gland [12, 13], oviduct [14], ductus deferens [15], urinary tract [16–18], uterus [19], and in the blood vessels [20, 21].

Firstly ICC cells have not been found in the gallbladder [22]. However it was Xiaomin *et al.* who found in 2006 ICC cells in the mice gallbladder, and Lavoie *et al.* in 2007 of the guinea pig [23, 24]. In humans it was Ortiz-Hidalgo *et al.* who found ICC in the human gallbladder [25].

To conclude, the role of ICC is associated with: generation and modulation of slow waves within gastrointestinal tract, facilitation of propagation of slow waves, their coordination and mediation in transmission of excitatory or inhibitory stimu-

lation (neurotransmission) between autonomic nervous system and muscular cells.

In certain diseases of the alimentary system the number of interstitial cells is significantly decreased. The direct mechanism of regression of Cajal cells is apoptosis [26], trans/de-differentiation [27, 28], while regeneration of ICC is associated with: proliferation of mature ICC [29], renovation of damaged cells and restoration from stem cells [30]. The newest reports say that mature ICC can still proliferate [31–33]. Kit signal path activated by kit-ligand is associated with control of survival and proliferation of ICC. We know also, playing similar role, other signal paths with neuronal originated nitric oxide [34], serotonin acting through 5-HT_{2B} receptor [35], interleukine-9 [36], insulin and insulin-like growth factor (IGF-1) [37], heme oxygenase (HO-1) [38]. 5-HT_{2B} receptor is responsible for maintenance of network of ICC [39]. Its activation begins proliferation of ICC. Knock-out mice which did not have that mutation of locus W, coding c-kit protein in mice (W^v/W) and rats Ws/Ws) leads to decrease of interstitial cells of Cajal in myenteric plexus of small intestine and stomach, what causes regression of slow waves [40]. Similar result was achieved by application of antibodies which blocked c-kit receptor in mice. Decrease of number or malfunction of interstitial cells is associated with numerous diseases of gastrointestinal system, i.e. idiopathic perforation of stomach, Hirschprung disease, gastroparesis and many others [41]. Changes according to number and function of interstitial cells which integrate motor activity of alimentary tract may play role also in gallstone disease [42–46]. Mutation in gene coding kit receptor leads to transformation of interstitial cells and gastrointestinal stromal tumors (GIST) development [47].

MATERIAL AND METHODS

The studies were carried out on 55 patients (19 males and 39 females, aged 27–80) operated in the 1st Department of General Surgery JU MC in 2010. The study was approved by Ethical Committee of Jagiellonian University (KBET/30/B/2010) according to Helsinki Declaration.

30 of patients were operated due to symptomatic cholelithiasis — using laparoscopic cholecystectomy. Before surgery in all patients normal level of bilirubin was confirmed. Control was based on 25 patients qualified to planned surgery because of tumor of pancreatic head, with no pre- and postoperative signs of cholelithiasis. Normal level of bilirubin was also required.

Obtained gallbladders were sectioned longitudinally and washed in phosphate buffer saline (PBS), next they were placed in 4% solution of buffered paraformaldehyde. The staining last 24 hrs. in room temperature. Next the material was washed in PBS. The gallbladders were examined macroscopically. An excision from each specimen was made — a band 4–5 mm wide, including the fundus, the body and the neck. Excisions obtained were properly marked with the ink. Next

the material was dehydrated in rising concentrations of alcohol, then washed in xylene. After immersing in paraffin at the temperature 56°C, tissue blocks were made. Next paraffin blocks were cut on microtome (Jung BioCut 2035, Leica, Wetzlar, Germany) into slices 6µm thick. The slices were deparaffinized in xylene (for immunostaining) and next placed in PBS solution. As control we used paraffin blocks of ileum obtained from collection of Department of Histology, Jagiellonian University, Medical College. To visualize the interstitial cells of Cajal (ICC) we used the method which was described in [46].

RESULTS

Pathologic evaluation of the gallbladders studied showed signs of chronic inflammation (chronic cholecystitis) in the wall. Immunohistochemical methods used (indirect double immunofluorescence staining) allowed identification of cells which revealed positive reaction with c-kit antibody. Among these cells we could find both ICC and numerous mastocytes. Application of antibodies against mast tryptase (normally found within mastocytes) enables differentiation between mast cells and ICC. Under the microscope we could observe c-kit immunopositive cells (red) and positive to tryptase (green), more — bluish nuclei, which one could see using specific filters. For statistical analysis ICC were: c-kit immunopositive, tryptase-immunonegative, presenting nuclei (to distinguish them from incidental non-nuclear structures). ICC were visible in the entire area of the band including fundus, body and neck of the gallbladder. They were found practically only within muscular layer, where they paralleled smooth muscle fibers. Microscopic analysis of specimens proved the following distribution of interstitial cells: intramuscular (ICC-IM), between the fibers of smooth muscle; interbundle — ICC-IB — within the connective tissue separating bundles of smooth muscle fibers. ICC existed usually as single cells, sometimes in groups of 2–3 cells, but we did not find networks of these cells.

Interstitial cells possessed characteristic elongated, fusiform shape — their length varied from 40–60 µm, in some specimens we could observe their processes. Besides, in some specimens we have observed few c-kit immunopositive cells (being simultaneously tryptase-negative), roundish in shape, which following the configuration of immunologic markers were assessed as ICC. C-kit immunopositive mast-cellshad usually roundish or oval shape, centrally placed nucleus, what enabled their differentiation from ICC. In control the average number of the interstitial cells was 7.06 per one visual field. We did not analyze differences between certain parts of gallbladder regarding the content of ICC. The average number of ICC obtained from bands of gallbladders of patients with cholelithiasis was 3,35/one visual fiend and it was twice lower in comparison to the control. The difference was statistically significant ($p < 0,0001$). We did not observe diffe-

rences in morphology and size of ICC which came from group of patients with cholelithiasis and control, estimated under the light microscope. We did not find also differences in distribution of ICC. We did not state dependence between the number of ICC and age.

DISCUSSION

Cholelithiasis is nowadays a major problem of modern medicine. Operations on gallbladder (cholecystectomies) due to gallstone disease are the most common procedures carried out in surgical departments. Following current trends cholelithiasis is a polietiologic disease. Pathogenesis of gallstone disease is a subject of many articles and according to widely accepted theories it is caused mainly by disturbances of lipid composition of bile and bile retention resulted from weakening of motor activity of gallbladder [48–51].

Understanding of physiology of smooth muscles of gastrointestinal system throughout last decades has significantly improved, mostly thank to studies on population of ICC. Discovery of immunoreaction of ICC with antibody against c-kit antigen opened new era in methodology of identification of ICC. ICC are present in numerous organs, but their morphology and function was best recognized in alimentary tract. It was established that these cells play role of pacemakers and their dysfunction may become a basis for motor disturbances of gastrointestinal system. This is why it seems that role of ICC is associated with: generation and modulation of slow waves in gastrointestinal tract, facilitation and propagation (promotion) of slow waves and their coordination, transition of transmission of excitatory or inhibitory stimuli (neurotransmission) between the autonomic nervous system and muscular cells. Knowledge on dependence of ICC and c-kit receptor for their normal development and maturation was a milestone, because it began wide application of immunohistochemical technics for identification of ICC [52].

ICC in the biliary system are not so long known. For the first time it was Ortiz-Hidalgo *et al.* in 2000 who presented stromal tumor of gallbladder containing cells phenotypically similar to ICC [25]. In 2007 Hinescu *et al.* described for the first time ICC cells in human gallbladder removed for non-neoplastic reasons [26]. Ahmadi *et al.* found ICC cells in extrahepatic biliary tracts in 2009 [53]. There's a lack of further reports on it in current literature, nor information on the distribution of ICC in the subsequent layers of wall of human gallbladder. In present study a number of ICC cells were identifies in the wall of gallbladder using indirect double immunofluorescence with antibodies against c-kit receptor and mast-tryptase.

Hinescu *et al.* [26] to identify ICC cells used immunoperoxidase staining with antibody anti-c-kit/CD 117. However we must remember that such staining allows dying of all c-kit positive cells, both ICC and mast cells. Sometimes it is

not possible to differentiate these two lines of cells using morphological features seen under the microscope. From another hand however Ahmadi *et al.* used the same method which we used [53].

In our study we found significant decrease of number of ICC cells in the wall of gallbladders of patients suffering from gallstone disease in comparison to the control. Such decrease may influence motor activity of the gallbladder. Numerous articles indicate different dysfunctions of the gallstone diseased patients [54–60]. We can conclude that such change leads to problems in bile flow, what in consequence causes development and deposition of concretions in the lumen of the gallbladder. Considering the role of ICC cells for the motor activity of the gastrointestinal system, the decreased number of ICC in the wall of the gallbladder correlates with motor dysfunction of the organ and reduction of number of interstitial cells is an important pathognomonic factor in biliary lithiasis.

CONFLICT OF INTERESTS

None declared.

ACKNOWLEDGEMENTS

Authors with to thank Dr Krzysztof Gil for substantial support with image analysis techniques and Dr Mariusz Gajda and Piotr Doba for excellent technical assistance.

REFERENCES

1. Cajal S.R.: Nuevas aplicaciones del metodo de coloracion de Golgi. Gaceta Medica Catalana. 1889; 12: 613–616. — 2. Cajal S.R.: Sur le ganglions et plexus nerveux de l'intestin. C R Soc Biol Paris. 1893; 5: 217–223. — 3. Maeda H., Yamagatan A., Nishikawa S.: Requirement of c-kit for development of intestinal pacemaker system. Development. 1992; 116 (2): 369–375. — 4. Ishikawa K., Komuro T., Hirota S.: Ultrastructural identification of the c-kit expressing interstitial cells in the rat stomach: a comparison of control and Ws/Ws mutant rats. Cell Tiss Res. 1997; 289 (1): 137–143. — 5. Komuro T., Zhou D.S.: Anti-c-kit protein immunoreactive cells corresponding to the interstitial cells of Cajal in the guinea-pig small intestine. J Auton Nerv Syst. 1996; 61 (2): 169–174. — 6. Komuro T.: Structure and organization of interstitial cells of Cajal in the gastrointestinal tract. J Physiol. 2006; 576: 653–658. — 7. Popescu L.M., Hinescu M.E., Ionescu N.: Interstitial cells of Cajal in pancreas. J Cell Mol Med. 2005; 9: 169–190. — 8. Hinescu M.E., Popescu L.M.: Interstitial Cajal-like cells (ICLC) in human atrial myocardium. J Cell Moll Med. 2005; 9: 972–975. — 9. Hinescu M.E., Gherghiceanu M., Mandache E.: Interstitial Cajal-like cells (ICLC) in atrial myocardium: ultrastructural and immunohistochemical characterization. J Cell Moll Med. 2006; 10: 429–443. — 10. Popescu L.M., Gherghiceanu M., Hinescu M.E.: Insights into interstitium of ventricular myocardium: interstitial Cajal-like cells (ICLC). J Cell Moll Med. 2006; 10: 444–458.
11. Shafik A., El-Sibai O., Shafik I.: Immunohistochemical identification of the pacemaker Cajal cells in the normal human vagina. Arch Gynecol Obstet. 2005; 272 (1): 13–16. — 12. Radu E., Re-

galia T., Ceafalan L.: Cajal-type cells from human mammary gland stroma: phenotype characteristics in cell culture. *J Cell Moll Med.* 2005; 9: 748–752. — **13.** Gherghiceanu M., Popescu L.M.: Interstitial Cajal-like cells (ICLC) in human resting mammary gland stroma. Transmission electron microscope (TEM) identification. *J Cell Moll Med.* 2005; 9: 893–910. — **14.** Popescu L.M., Ciontea S.M., Cretoiu D.: Novel type of interstitial cell (Cajal-like) in human Fallopian tube. *J Cell Moll Med.* 2005; 9: 479–523. — **15.** Metzger R., Rolle U., Fiegel H.C.: C-kit receptor in the human vas deferens: distinction of mast cells, interstitial cells and interepithelial cells. *Reproduction* 2008; 135: 377–384. — **16.** Van der Aa F., Roskams T., Blyweert W.: Identification of kit positive cells on the human urinary tract. *J Urol* 2004; 171: 2492–2496. — **17.** Lang R.J., Klemm M.F.: Interstitial cell of Cajal-like cells in upper urinary tract. *J Cell Moll Med.* 2005; 9: 543–556. — **18.** Sergeant G.P., Thonbury K.D., McHale N.G.: Interstitial cells of Cajal in the urethra. *J Cell Moll Med.* 2006; 10: 280–291. — **19.** Ciontea S.M., Radu E., Regalia T.: C-kit immunopositive interstitial cells (Cajal-type) in human myometrium. *J Cell Moll Med* 2005; 9: 407–420. — **20.** Harhun M., Pucovsky V., Povstyan O.V.: Interstitial cells in the vasculature. *J Cell Moll Med* 2005; 9: 232–243.

21. Harhun M., Gordienko D., Kryshchal D.: Role of intracellular stores in the regulation of rhythmic $[Ca^{2+}]$ changes in interstitial cells of Cajal from rabbit portal vein. *Cell Calcium.* 2006; 40: 287–298. — **22.** Parr E.J., Kennedy A.L., Maue G.M.: Lack of evidence for the existence of interstitial cells of Cajal in the gallbladder. *Gastroenterol.* 2003; 124 (1): 347–349. — **23.** Lavoie B., Balemba O.B., Nelson M.T.: Morphological and physiological evidence for interstitial cells of Cajal-like cells in the guinea pig gallbladder. *J Physiol.* 2007; 579: 487–501. — **24.** Sun X., Yu B., Xu L.: Interstitial cells of Cajal in the murine gallbladder. *Scand J Gastroenterol* 2006; 41: 1218–1226. — **25.** Ortiz-Hidalgo C., de Leon Bojorge B., Albores-Saavedra J.: Stromal tumor of the gallbladder with phenotype of interstitial cells of Cajal: a previously unrecognized neoplasm. *Am J Surg Pathol.* 2000; 24: 1420–1423. — **26.** Hinescu M.E., Ardeleanu C., Gherghiceanu M.: Interstitial Cajal-like cells in human gallbladder. *J Mol Histol.* 2007; 38: 275–284. — **27.** Lecoin L., Glabella G., LeDouarin N.M.: Origin of the c-kit positive interstitial cells in the avian bowel. *Development.* 1996; 122: 725–733. — **28.** Young H.M., Ciampoli D., Southwell B.R.: Origin of interstitial cells of Cajal in the mouse intestine. *Dev Biol.* 1996; 180: 97–107. — **29.** Young H.M.: Embryological origin of interstitial cells of Cajal. *Microsc Res Tech.* 1999; 47: 303–308. — **30.** Thomsen L., Robinson T.L., Lee J.C.F.: Interstitial cells of Cajal generate a rhythmic pacemaker current. *Nature Med.* 1998; 2: 848.

31. Ward S.M., Ordog T., Koh S.D.: Pacemaking in interstitial cells of Cajal depends upon calcium handling by endoplasmic reticulum and mitochondria. *J Physiol.* 2000; 525 (2): 355–361. — **32.** Sanders K.M.: A case for interstitial cells of Cajal as pacemakers and mediators of neurotransmission in the gastrointestinal tract. *Gastroenterol.* 1996; 111 (2): 492–515. — **33.** Mei F., Zhu J., Guo S.: An age-dependent proliferation is involved in the postnatal development of interstitial cells of Cajal in the small intestine of mice. *Histochem Cell Biol.* 2009; 131: 43–53. — **34.** Choi K.M., Gibbons S.J., Roeder J.L.: Regulation of interstitial cells of Cajal in the mouse gastric body by neuropeptide nitric oxide. *Neurogastroenterol Motil.* 2007; 19: 585–595. — **35.** Wouters M.M., Gibbons S.J., Roeder J.L.: Exogenous serotonin regulates proliferation of interstitial cells of Cajal in mouse jejunum through 5-HT_{2B} receptors. *Gastroenterol.* 2007; 133: 897–906. — **36.** Ye J., Zhu Y., Khan W.: IL-9 enhances growth of ICC, maintains network structure and strengthens rhythmicity of contraction in culture. *J Cell Moll Med.* 2006; 10: 687–694. — **37.** Horvath V.J., Vittal H., Lornicz A.: Reduced stem cell factor links smooth myopathy and loss of interstitial cells of Cajal in murine diabetic gastroparesis. *Gastroenterol.* 2006; 130: 759–770. — **38.** Choi K.M., Gibbons S.J., Nguyen T.V.: Heme oxygenase-1 protects interstitial cells of Cajal from oxidative stress and reverses diabetic gastroparesis. *Gastroenterol.* 2008; 135: 2055–2064. — **39.** Tharayil V.S., Wouters M.M., Stanich J.E.: Lack of serotonin 5-HT_{2B} receptor alters proliferation and network volume of interstitial cells of Cajal in vivo. *Neurogastroenterol Motil.* 2010; 22 (4): 462–469. — **40.** Takayama I., Horiguchi K., Daigo Y.: The interstitial cells of Cajal and a gastroenteric pacemaker system. *Arch Histol Cytol.* 2002; 65 (1): 1–26.

41. Streutker C.J., Huizinga J.D., Driman D.K.: Interstitial cells of Cajal in health and disease. Part I: Normal ICC structure and function with associated motility disorders. *Histopath.* 2007; 50:

176–189. — **42.** Pasternak A., Gil K., Gajda M., Tomaszewski K.A., Matyja A., Walocha J.A.: Interstitial Cajal-Like Cell: A New Player in Cholelithiasis? *Am. J. Gastroenterol.* 2014; 109 (4): 603–604. — **43.** Matyja A., Gil K., Pasternak A., Sztęfko K., Gajda M., Tomaszewski K.A., Matyja M., Walocha J.A., Kulig J., Thor P.: Telocytes: new insight into the pathogenesis of gallstone disease. *J Cell Mol Med.* 2013; 17 (6): 734–742. — **44.** Pasternak A., Gil K., Matyja A., Gajda M., Sztęfko K., Walocha J.A., Kulig J., Thor P.: Loss of gallbladder interstitial Cajal-like cells in patients with cholelithiasis. *Neurogastroenterol Motil.* 2013; 25 (1): e17–e24. — **45.** Pasternak A., Matyja A., Gil K., Gajda M., Tomaszewski K., Matyja M., Walocha J.A., Kulig J.: Interstitial Cajal-like cells and bile lithogenicity in the pathogenesis of gall-stone disease. *Pol Przegl Chir.* 2013; 85 (6): 311–316. — **46.** Pasternak A., Gajda M., Gil K., Matyja A., Tomaszewski K.A., Walocha J.A., Kulig J., Thor P.: Evidence of interstitial Cajal-like cells in human gallbladder. *Folia Histochem Cytobiol.* 2012; 50 (4): 581–585. — **47.** Cichoż-Lach H., Kasztelan-Szczerbińska B., Słomka M.: Stromalne guzy przewodu pokarmowego — epidemiologia, obraz kliniczny, diagnostyka, rokowanie oraz zasady leczenia. *Pol Arch Med Wewn.* 2008; 118 (4): 216–221. — **48.** Paumgartner G., Sauerbruch T.: Gallstones: Pathogenesis. *Lancet.* 1991; 338: 1117–1121. — **49.** Holzbach R.T.: Gallbladder stasis: consequence of long-term parenteral hyperalimentation and risk factor of cholelithiasis. *Gastroenterol.* 1983; 84: 1055–1058. — **50.** Portincasa P., Moschetta A., Palasciano G.: Cholesterol gallstone disease. *Lancet.* 2006; 368: 230–239.

51. Strasberg S.M.: The pathogenesis of cholesterol gallstones — a review. *J Gastrointest Surg.* 1998; 2: 109–125. — **52.** Torihashi S., Ward S.M., Nishikawa S.: C-kit dependent development of interstitial cells and electrical activity in the murine gastrointestinal tract. *Cell Tiss Res.* 1995; 280 (1): 97–111. — **53.** Ahmadi O., de l'Nicolson M., Gould M.: Interstitial cells of Cajal are present in human extrahepatic bile ducts. *J Gastroenterol Hepatol.* 2010; 25 (2): 277–285. — **54.** Festi D., Frabboni R., Bazzoli F.: Gallbladder motility in cholesterol gallstone disease. Effect of ursodeoxycholic acid administration and gallstone dissolution. *Gastroenterol.* 1990; 98: 1779–1785. — **55.** Stolk M.F.J., van Erpecum K.J., Samson M.: Interdigestive gallbladder emptying, antroduodenal motility and motilin release in cholesterol gallstone patients. In: Stolk MFJ ed. *Pathogenesis of cholesterol gallstones.* Utrecht: Thesis Universiteit Utrecht, 1993: 65–78. — **56.** Portincasa P., Di Ciaula A., Vendemiale G.: Gallbladder motility and cholesterol crystallization in bile from patients with pigment and cholesterol gallstones. *J Clin Invest.* 2000; 30: 317–324. — **57.** Pomeranz I.S., Shaffer E.A.: Abnormal gallbladder emptying in a subgroup of patients with gallstones. *Gastroenterol.* 1985; 88: 787–791. — **58.** Sharma B.C., Agarwal D.K., Dhiman R.K.: Bile lithogenicity and gallbladder emptying in patients with micro-lithiasis: effect of bile acid therapy. *Gastroenterol.* 1998; 115: 124–128. — **59.** Trierney S., Pitt H.A., Lillemoe K.D.: Physiology and pathophysiology of gallbladder motility. *Surg Clin North Am.* 1993; 73 (6): 1267–1291. — **60.** Doty J.E., Pitt H.A., Kechenbecker S.L.: Impaired gallbladder emptying before gallstone formation in the prairie dog. *Gastroenterol.* 1983; 85: 168–174.

¹ Department of Anatomy

Jagiellonian University Medical College
ul. Kopernika 12, 31-034 Kraków, Poland
Head: prof. dr hab. Jerzy Walocha

² 1st Department of General Surgery

Jagiellonian University Medical College
ul. Kopernika 40, 31-501 Kraków, Poland
Head: prof. dr hab. Jan Kulig

³ 2nd Chair of General Surgery

Jagiellonian University Medical College
ul. Kopernika 21, 31-501 Kraków, Poland
Head: prof. dr hab. Andrzej Budzyński

Corresponding author:

Artur Pasternak MD, PhD
Department of Anatomy
Jagiellonian University Medical College
ul. Kopernika 12, 31-034 Kraków, Poland
Phone: +48 12 422 95 11
E-mail: artur.pasternak@uj.edu.pl

